

# Robot Control with Biological Cells

Soichiro Tsuda<sup>a,\*</sup>, Klaus-Peter Zauner<sup>b</sup>, Yukio-Pegio Gunji<sup>a</sup>

<sup>a</sup>*Graduate School of Science and Technology  
and Department of Earth & Planetary Science  
Kobe University Nada, Kobe 657-8501, Japan*

<sup>b</sup>*School of Electronics and Computer Science  
University of Southampton, SO17 1BJ, United Kingdom*

---

## Abstract

At present there exists a large gap in size, performance, adaptability and robustness between natural and artificial information processors for performing coherent perception-action tasks under real-time constraints. Even the simplest organisms have an enviable capability of coping with an unknown dynamic environment. Robots, in contrast, are still clumsy if confronted with such complexity. This paper presents a bio-hybrid architecture developed for exploring an alternate approach to the control of autonomous robots. Circuits prepared from amoeboid plasmodia of the slime mold *Physarum polycephalum* are interfaced with an omnidirectional hexapod robot. Sensory signals from the macro-physical environment of the robot are transduced to cellular scale and processed using the unique micro physical features of intracellular information processing. Conversely, the response from the cellular computation is amplified to yield a macroscopic output action in the environment mediated through the robot’s actuators.

*Key words:* autonomous robots, molecular computing, coupled oscillators, biologically-inspired robotics

---

## 1 Introduction

The prevalent approach to robot control is based on ‘behaviour decomposition’. The interaction loop between robot and environment is decomposed and treated as individual modules. This concept simplifies controller design and proved successful in many robotic systems including humanoid robots (e.g.,

---

\* Corresponding author: 026d874n@y04.kobe-u.ac.jp

Fujita et al., 2003). With ‘behaviour decomposition’, the flexible selection of modules is critical for an adaptive and autonomous behaviour of the robot. One approach to addressing the decomposition problem disaggregates learning, and interaction with the environment. While in learning mode, the robot learns or self-organises a proper module in terms of some environmental signal provided by a teacher, and subsequently it uses this module in interaction mode.

The use of ‘behaviour decomposition’ enables robots to successfully work in either a stable work space or with the support of a teacher. The scheme can be implemented in localised (Tani and Nolfi, 1999) or distributed fashion (Tani et al., 2004). Without a teacher, however, delineating a boundary for the environment becomes an insurmountable challenge as the environment has no natural limit. In an unknown dynamic environment a disaggregation of learning mode and interaction mode is not useful unless this manifestation of the ‘frame problem’ (McCarthy and Hayes, 1969) can be overcome. In an attempt to address this difficulty Brooks (1990) proposed the ‘subsumption architecture’ and implemented it in insect-like robots (Brooks, 1991). This route, however, proved difficult to scale up because of computational cost. Quite a number of extension and modification of the above architectures have been reported, but the quest for genuinely autonomous robot control, capable of coping with an unknown complex environment, has not yet been successful. Nonetheless, organisms seemingly do not face the difficulties encountered with robots. Cariani (1992) points to the plasticity of architecture and its ability to form appropriate sensory, computational, and effector structures in response to direct interaction with the environment. Whether it is possible to replicate this plasticity on a virtual level in a formal computational model for execution on digital machine is unclear. Undoubtedly, simulation is a powerful tool for studying design concepts. Ziegler et al. (1998) were among the first to consider the biochemical metaphor for robot control and employed simulation to evolve highly abstracted chemical controllers. More recently Adamatzky et al. (2004) have taken steps towards the realisation of a chemically controlled robot based on reaction diffusion computing with an excitable chemical medium (cf. Rambidi, 2005).

Conrad (1989, 1999) argued that the common digital mode of computation is fundamentally different from the context-sensitive mode of information processing found in biological systems and any attempt to emulate it on a digital computer would be inherently inefficient. If Conrad’s assertion holds then robots that face an unknown complex dynamic environment cannot be successfully controlled with a conventional computer. This is the case because the requirement to act in real-time precludes the use of an architecture that trades efficiency for programmability (Conrad, 1995). The medium of computation becomes significant. On this basis we decided to explore an approach that recruits intracellular dynamics to autonomous robot control.

### 1.1 Intracellular computation

Ideas from biological information processing have influenced numerous robot control architectures. Most of them draw their inspiration from neural networks. As a matter of fact, however, almost all known biological information processing occurs at the subcellular level. In consequence of the tremendous success of single cell organisms in our biosphere, this is the case even if we set aside molecular level computation within neurons. The significance of intracellular information processing has been recognised three decades ago (Conrad, 1972; Liberman, 1979). The picture that emerged since then shows a host of biochemical processes in the cell and its membrane contributing to signal processing and computing. Thus far the mechanisms that confer the enviable computation power to cells are not understood. It is possible, however, to discern properties of the intracellular medium that contribute. Two principles are pertinent. One is shape-based self-assembly: Brownian motion provides a search mechanism that in combination with free energy minimization enables the recognition of specific molecular coded signals in a highly complex chemical background. The other is conformational state change: the complex electrostatic interaction network formed by the atoms within a macromolecule integrates the electrostatic environment of the molecule and in accordance therewith modulates the properties of the molecule. Self-assembly facilitates communication among distant molecules, but is limited by diffusion speed. It can serve for signal integration through the assembly of multi-component supramolecular structures. More typically, conformational state change mediates signal fusion. The conformational dynamics in a molecule can be very rapid, and signal propagation along lines of molecules in direct contact is accordingly fast. The two principles above are implemented with a large number of specialised components that have been picked through evolution from a combinatorially vast space of potential combinations of building blocks. The building block scheme extends beyond the molecular level to the cells themselves. A myriad of different cells, each specialised to operate in a particular environment, are found in nature. Let us now turn to our cell of choice for the robot controller.

## 2 *Physarum polycephalum*

The amoeboid plasmodium of the slime mold *Physarum polycephalum* is a large multi-nuclear single-cellular organism (Fig. 1A). In a fully developed plasmodium flat sheet-like areas at the edge are connected by more centrally located tubular structures as visible in Fig. 1B. Being a single cell the plasmodium— which can extend to over 20 m in diameter — does not possess a nervous system. It relies on intracellular information processing to integrate local sen-

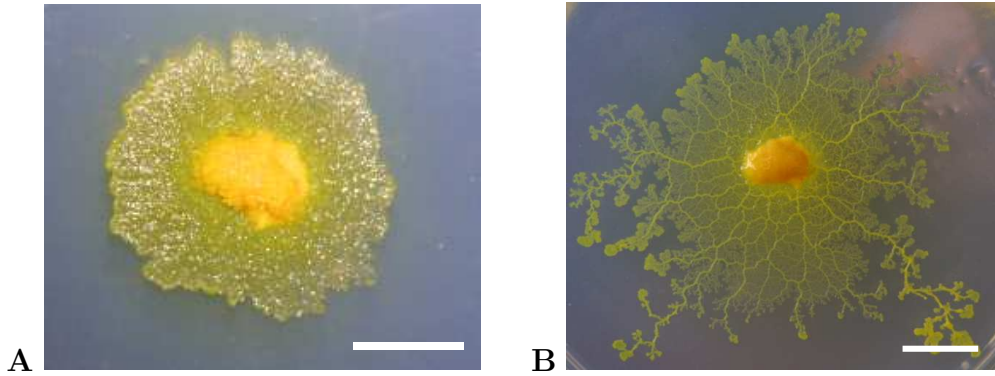


Fig. 1. Plasmodium of *Physarum polycephalum*. The early stage (A) and in developed form, approximately eight hours later (B). In both panels the length of the white bar corresponds to 10 mm.

sory information and to generate an appropriate response to stimuli. Different areas within the plasmodium communicate by protoplasmic flow. This oscillatory flow of protoplasm is called shuttle streaming and provides a transport mechanism for nutrition and signals. A dynamically reconfiguring network of tubes, formed from gelled cytosol, directs the flow of protoplasm and also the overall movement and growth direction of the cell. It has been observed that the rhythm of shuttle streaming is synchronised with intracellular oscillatory behaviours such as oscillations in ATP concentration,  $\text{Ca}^{2+}$  concentration, and the plasmagel/plasmasol exchange rhythm (Ueda et al., 1986). This oscillatory rhythm is known to increase in frequency in the presence of attractive stimuli (glucose, warmth) and conversely, slows if a repulsive stimulus (blue light, salt) is encountered (Durham and Ridgway, 1976). The plasmodium is often viewed as a system of coupled oscillators and the oscillations are implicated in the information processing mechanisms of *Physarum* plasmodium. The oscillatory behaviour has been studied from the dynamical systems perspective. A collective oscillator model with a limit cycle was introduced by Takamatsu et al. (1997). Recently Tero et al. (2005) suggested a reaction-diffusion model for cell fusion between two plasmodia that exhibits an anti-phase oscillation characteristic.

The plasmodium can be regarded as an autonomous distributed system. In a series of studies Nakagaki and coworkers found that this natural parallel processor can solve small instances of optimisation problems and find an optimal path through a maze (Nakagaki et al., 2000, 2001). In the context of the present work the patterning of the plasmodium with masks is particularly relevant (Takamatsu et al., 2000a,b). Our architecture for a physarum robot controller draws on this technique for constructing coupled nonlinear oscillator circuits (Takamatsu and Fujii, 2002).

### 3 Cellular Robot Control

The objective here is to make the microphysical scale of cellular information processing amenable to robotic control applications. To achieve this aim we require a system in which signals from the environment are transduced to a cell and processed using the micro physical features of intracellular information processing. Then, the results of the cellular computation need to be amplified to steer the robot. After these two components of the cell-robot interface have been established, the intracellular level information processing can engage with the macroscopic environment of the robot in an interaction loop across physical scales. (Fig. 2).

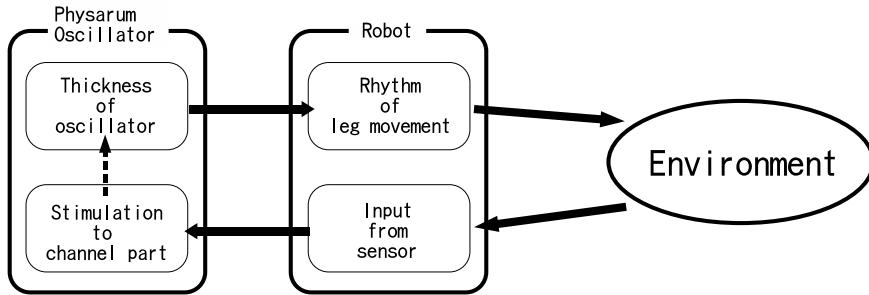


Fig. 2. Cell-robot-environment interaction loop

Conrad (1996) emphasised the importance of the microphysical scale for the problem solving behaviour of biological systems. He introduced the conceptual picture of a vertical percolation of signals through a hierarchy of physical structures. The underlying assumption of this concept is that evolution recruited the dynamics on all levels of scale for the efficient information processing required in autonomous systems capable of negotiating a complex hostile environment in real-time. To instantiate such a device and to provide a testbed for exploring and evaluating of bio-mimetic principles of cross-scale information processing in artificial devices we have implemented a robot and the requisite cell-robot interface.

#### 3.1 A pole-free legged robot

An omnidirectional walker design was adopted for the robot, because it enables us to utilize the oscillatory behaviour of the cell more directly for driving locomotion and we can more readily understand the dynamics of the robot. The six-legged configuration with a hexagonal body shape is pictured in Fig. 3. The hexagonal platform is equipped with sensors equally in all six directions, resulting in a pole-free robot that can move and sense omnidirectional.

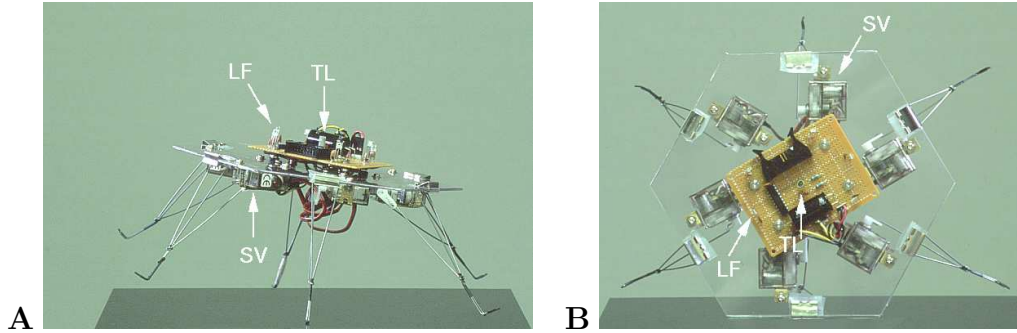


Fig. 3. Six-legged robotic platform, side view (A) and top view (B). The hexagonal body carries six light-to-frequency converters (LF) and six legs driven by servos (SV). To assist in tracking the 2D-movements of the robot, two LEDs are used as tracking lights (TL).

The chassis of the robot is a 2 mm thick sheet of acrylic plastic in the shape of a hexagon with 115 mm edge length. A circuit board mounted on the top of the chassis (visible in Fig. 3B) carries 6 light to frequency converters (TSL237-S, TAOS Inc., Plano, TX-USA; [www.taosinc.com](http://www.taosinc.com)) connected to an octal line driver (74ACT240, Fairchild, South Portland, ME-USA; [www.fairchildsemi.com](http://www.fairchildsemi.com)). The top circuit board also carries a connector for a tether which supplies 5 V for the sensor circuitry (six TSL237 and one 74ACT240), separate 5 V for the leg actuators, 12 V for an RS232 serial interface, the RS232 signal, and the buffered signal lines of the six light sensors. Furthermore, two LEDs of different colour are mounted on the top circuit board to facilitate tracking of position and orientation of the robot with an overhead camera. The same octal line driver as installed on the robot (74ACT240) is used at the other end of the tether to receive the frequency-coded sensor readings and is connected through six adjustable voltage dividers to six input channels of a sound card (Sound Blaster Live-Value, Creative Technology Ltd., Singapore; [www.creative.com](http://www.creative.com)) in a personal computer (PC). Six pulse-width controlled mini servos (PARK HPX, Grand Wing Servo-Tech Co. Ltd., Taipei, Taiwan; [www.gws.com.tw](http://www.gws.com.tw)) of the type used in radio controlled air planes are mounted to the underside of the acrylic chassis. Each servo provides a nominal torque of 4.2 kg/cm at 4.8 V and drives an approximately 100 mm long triangular leg formed from 1 mm diameter steel wire. The pulse trains for the six servos are generated on-board with a peripheral interface controller (Mini SSC II, Scott Edwards Electronics Inc. Sierra Vista, AZ-USA; [www.seetron.com](http://www.seetron.com)) which is mounted centrally on the underside of the chassis and receives its commands through a serial port of the PC. A leg is mounted in centre of every edge of the hexagonal chassis (cf. Fig. 3B) with the spring of a 20 mm foldback paper clip and is driven perpendicular to the chassis' edge by a lever (1 mm steal wire) connected to the servo actuator. This simple design offers excellent static stability and allows for numerous possible gait patterns with only one degree of freedom for each leg.

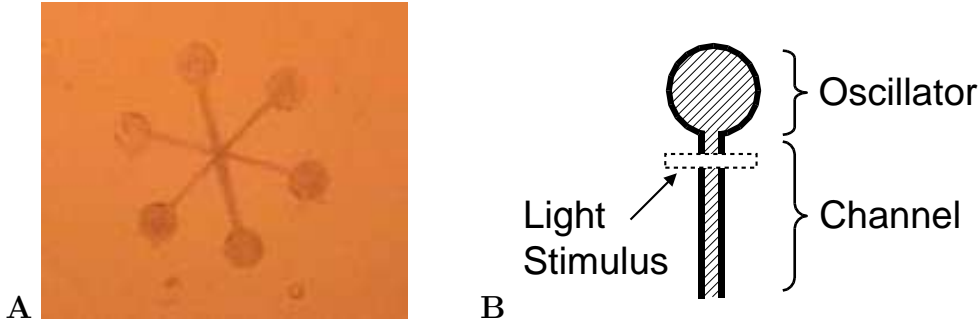


Fig. 4. *Physarum* oscillator circuit. The plasmodium is patterned as six oscillators with star coupling (A). The picture shows the view provided by the CCD camera indicated in Fig. 5. A diagrammatic view of one branch is shown on the right (B). The oscillator nodes have a diameter of 3 mm; channels are  $6 \times 0.5$  mm. A dashed box delineates the area irradiated for light stimulus.

### 3.2 *Physarum* circuits

#### 3.2.1 *Materials and Methods*

The plasmodia used for the experiments are cultured in the dark on wet paper towel (Crecia Co.,Ltd., Tokyo, Japan, [www.crecia.co.jp](http://www.crecia.co.jp)) in a plastic box at room temperature. Oatmeal is fed once or twice a day. All experiments have been conducted on a 1.5% agar gel plate (polystyrene 90 mm $\times$ 15 mm Petri dish, Ina Optica Co., Ltd.) with the plasmodium that was kept in food hunger for at least 6 hours. As noted above, the plasmodium prefers not to migrate from a moist surface to a dry region. For the dry masking of the the agar gel we chose plastic film (WT-OHP100, Seikyo, Kobe, Japan; [seagull.coop.kobe-u.ac.jp](http://seagull.coop.kobe-u.ac.jp)) which can easily be patterned to any desired shape with a sharp blade. In the experiment, the petridish with the plasmodium was placed on a fiber optic back light (1P100-1000F-1G8-MPM-R, OPTEL, Tokyo, Japan; [www.optel.co.jp](http://www.optel.co.jp)) and illuminated with with a halogen light source (OHX-100A, OPTEL) filtered with a 600 nm interference band pass (NT46-152, Edmund Optics Japan Co., Ltd., Tokyo, Japan; [www.edmundoptics.com](http://www.edmundoptics.com)). *Physarum* is not affected by light near this wavelength (Renzel et al., 2000). Light transmitted through the plasmodium was detected with a CCD camera (HOGA, Kyoto, Japan) and recorded typically at 2 seconds intervals by a PC through an image capture board (GV-VCP2/PCI, I-O Data, [www.iodata.jp](http://www.iodata.jp)). All experiments were conducted in the dark at 20°C.

#### 3.2.2 *Architecture of Physarum circuits*

In the experiments reported here we used the star shaped circuit shown in Fig. 4A. It consists of six circular wells of 3 mm diameter each, coupled at the centre through six channels. With this geometry, the part of the plasmodium

contained within a well will be synchronised and can be assumed to form a single oscillator (Takamatsu and Fujii, 2002). It is also known that adjacent plasmodia fuse into a single one and that, under the conditions considered here, plasmodia will never split up. This fact is used to grow the plasmodium into the mask. A tip portion ( $\approx 3 \times 3$  mm) cut from cultured larger plasmodium is placed on the side of each well as a seed. The mask thus prepared is incubated in the dark for at least 4 hours. The plasmodia migrate from the seeds onto the exposed agar surface and fuse at the centre where the channels meet. After the plasmodia fused into a single cell, incubation is continued for an additional 3 hours. After total of 7 hours incubation, tube structures are established in the channels. The circuit obtained corresponds to six individual nonlinear oscillators mutually coupled through the tubes formed in the circuit channels.

### 3.3 The cell-robot interface

Having implemented the above, it is necessary to combine the *Physarum* circuit with the robot. The architecture illustrated in figure 5 is based on an all-optical interface between cell and robot. It makes use of three phenomena. First, plasmodia are repelled by white light (Ueda et al., 1988). Second, the light transmission of plasmodia is inversely proportional to their thickness (Nakagaki et al., 1999). And, third, as previously stated, the plasmodium is insensitive to light near 600 nm (Renzel et al., 2000).

Signals from the robot's six light sensors are transduced to the plasmodium by shining a band of white light on the coupling channel of an oscillator (cf.

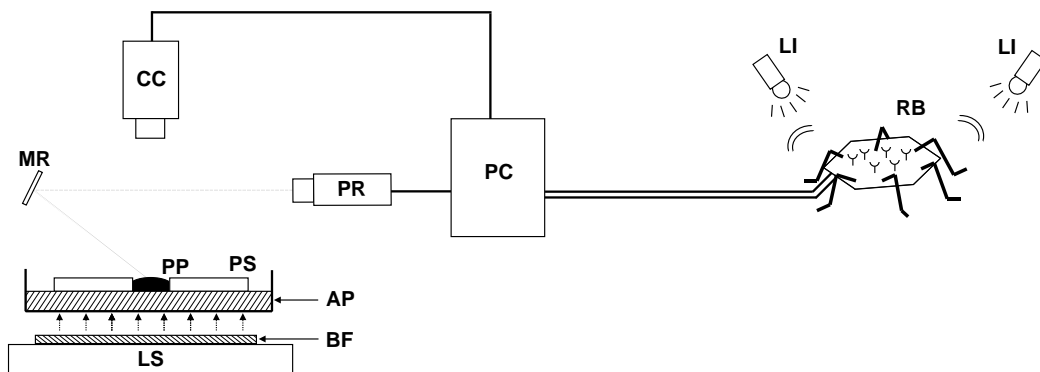


Fig. 5. Cellular robot controller. Sensors on the robot (RB) detect lamp input (LI) and transmit a signal to the computer (PC). The signals from the sensors are recoded by the PC into a spatial light pattern which is projected as light stimulus with a video projector (PR) via mirror (MR) onto the surface of the plasmodium (PP). Oscillations of the plasmodium (PP), which is patterned on an agar plate (AP) by a plastic sheet (PS), are detected by a CCD camera (CC) as intensity changes in light transmitted from the bandpass filtered (BF) light source (LS).

figure 4). To this end we project one mask from a set of dark masks with suitably positioned white lines onto the *Physarum* circuit. We use a video projector with standard optics and a light path of 1.2 m. By this way only a fraction of the projector's image covers the circuit, but the obtained resolution is sufficient to send signals to specific channels. Our observations suggest that light stimulation of an oscillator's channel weakens the coupling of this oscillator to other oscillators in the circuit. Such a weakening can alter the behaviour of the oscillator system.

To read out the response of the plasmodium, the circuit is illuminated through its agar base by orange light. The intensity of the transmitted light is recorded by a CCD camera. The image of the camera (shown in figure 4A) provides state information for all six oscillators patterned in the plasmodium. To calculate the amplitude of an oscillator, the average brightness values of a  $5 \times 5$ -pixel area centered over the well of the oscillator under consideration in the camera image is taken. For analysis purposes we eliminate camera noise by calculating moving temporal averages with a window size of 30 seconds.

It is now possible to transmit signals from the robot's light sensor to the plasmodium cell and to read the cell's oscillations in the six wells of the circuit, which is then used to control the robot's leg actuators. The oscillations of the plasmodium typically have a period in the order of 1–2 minutes. Practically, therefore, it is convenient to up-convert the oscillator frequencies rather than to use them directly for driving the actuators.

By using the arrangement above described the closed loop interaction between the plasmodium cell and the robot's environment depicted in figure 2 is established. Each plasmodial oscillator controls the rhythm of one robot leg. In combination the leg oscillations lead to a locomotion of the robot. As a consequence of the robot's motion, the light levels sensed by the robot changes. The change is communicated back to the plasmodium that responds with alternations in its oscillatory behaviour.

#### 4 Behaviour of a *Physarum* circuit

Two properties of the *Physarum* circuit are essential to make the conceived architecture practical. The coupled oscillatory system has to yield oscillation patterns that lead to robot gait. And, equally important, the gait pattern has to be responsive to stimuli. Figure 6 shows typical oscillatory behaviour of the wells in the *Physarum* circuit. The upper graph shows the relative thickness, i.e., the amplitude, measured for the six oscillator wells. The lower graph shows the binarized phase for the six oscillators on the same time axis as the graph above it. The six horizontal bands from top to bottom correspond to oscillators

1–6, respectively. A black vertical line for an oscillator indicates that at this point in time its thickness was increasing. Note that the oscillation pattern changes spontaneously, even without light stimulus, from a rotational pattern (a) to an alternating pattern (b). The position of oscillator labels used in the legend of the amplitude curves (upper graph) is shown in (c).

The anti-phase oscillation phenomenon supports robot locomotion. As will be described in the following section, an anti-phase oscillations give rise to an effective gait pattern. The experimental results obtained so far show that the six-oscillator circuit exhibits in-phase or anti-phase pattern under many circumstances. If we can modulate the relative phases of a pair of oscillators by light stimuli and manage to avoid interfering unfavourable oscillations, the robot can be made to walk in any desired direction under the control of the plasmodium cell.

Unexpectedly, we found that irradiating the channel of an oscillator has little effect on its rhythm. However, if a channel is stimulated with light and the stimulus is removed, the stimulated oscillator and the oscillator located directly opposite to it rapidly go into anti-phase oscillations as is shown in figure 7. A possible explanation of this phenomenon may be found in the fact that the driving force of protoplasmic flow is proportional to phase difference between two areas of the plasmodium (Nakagaki and Ueda, 1996). The switch to anti-phase oscillation upon removal of the light stimulus may be due to

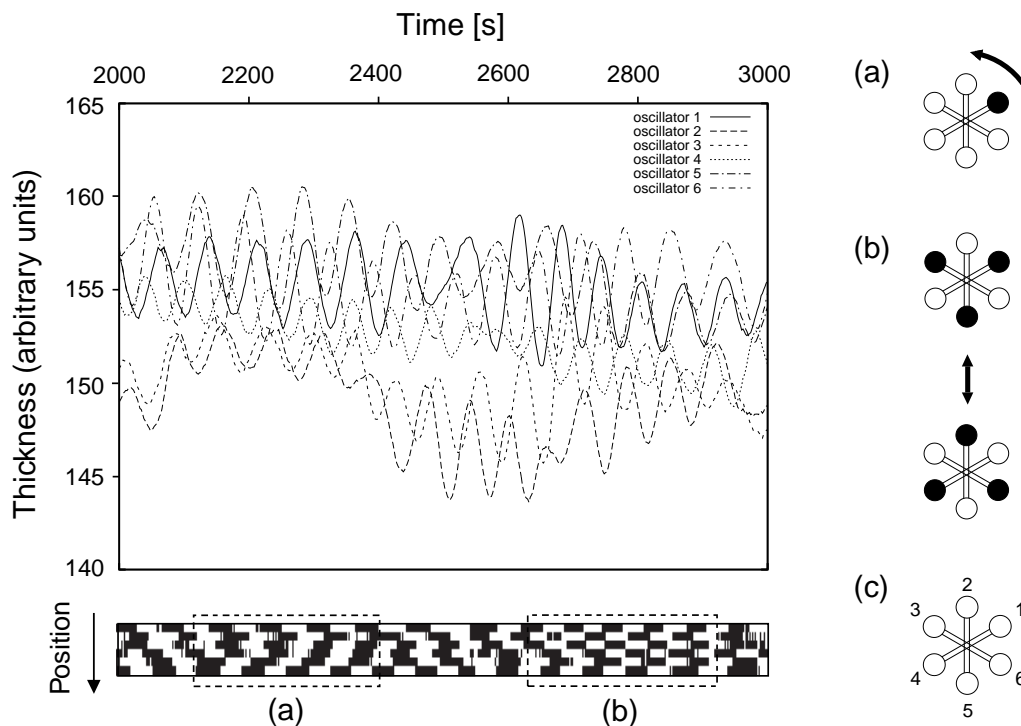


Fig. 6. Thickness oscillation of the plasmodium without light stimuli; sample selected from a long time series. See text for explanation.

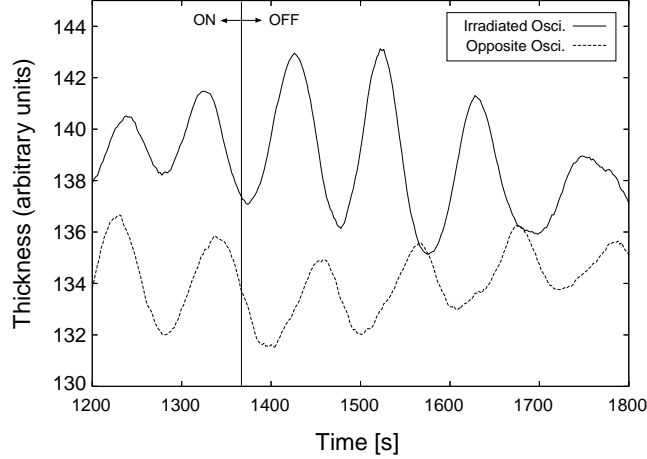


Fig. 7. Effect of light stimulus on oscillation of the plasmodium; sample selected from a long time series. The amplitude of an oscillator that receives a light stimulus is drawn as solid line, the amplitude of the oscillator opposite to it as dashed line. Irradiation is on until  $t=1370$  s (vertical line) and off thereafter. As soon as the light stimulus is removed, the oscillation switches rapidly from in-phase to anti-phase.

an autonomous recovering process. As shown by these experiments, the 6-oscillator star circuit design fulfils the two basic requirements for the cellular robot control architecture.

## 5 Robot Locomotion With Coupled Oscillators

To investigate the possible gait patterns that can be generated by coupled oscillator circuits and how the gait can be modulated we conducted a set of experiments in which the actuators of the robot are driven by a coupled nonlinear oscillator system implemented in software.

Each of the six legs of the robot is driven directly with the amplitude of one of six van der Pol oscillators. The choice of van der Pol oscillators was motivated by the work of Takahashi et al. (1997); Nomura (2001) on simulation of plasmodium behaviour. The fully (all-to-all) connected oscillators are described by:

$$\begin{aligned}
 dx_i(t)/dt &= y_i(t) \\
 dy_i(t)/dt &= \varepsilon(1 - 2\beta_i x_i(t) - x_i(t)^2)y_i - x_i(t) \\
 &\quad + \sum_{j=1}^6 \kappa_{ij}(x_j(t) - x_i(t)) \\
 &\quad + \sum_{j=1}^6 \sigma_{ij}(y_j(t) - y_i(t))
 \end{aligned} \tag{1}$$

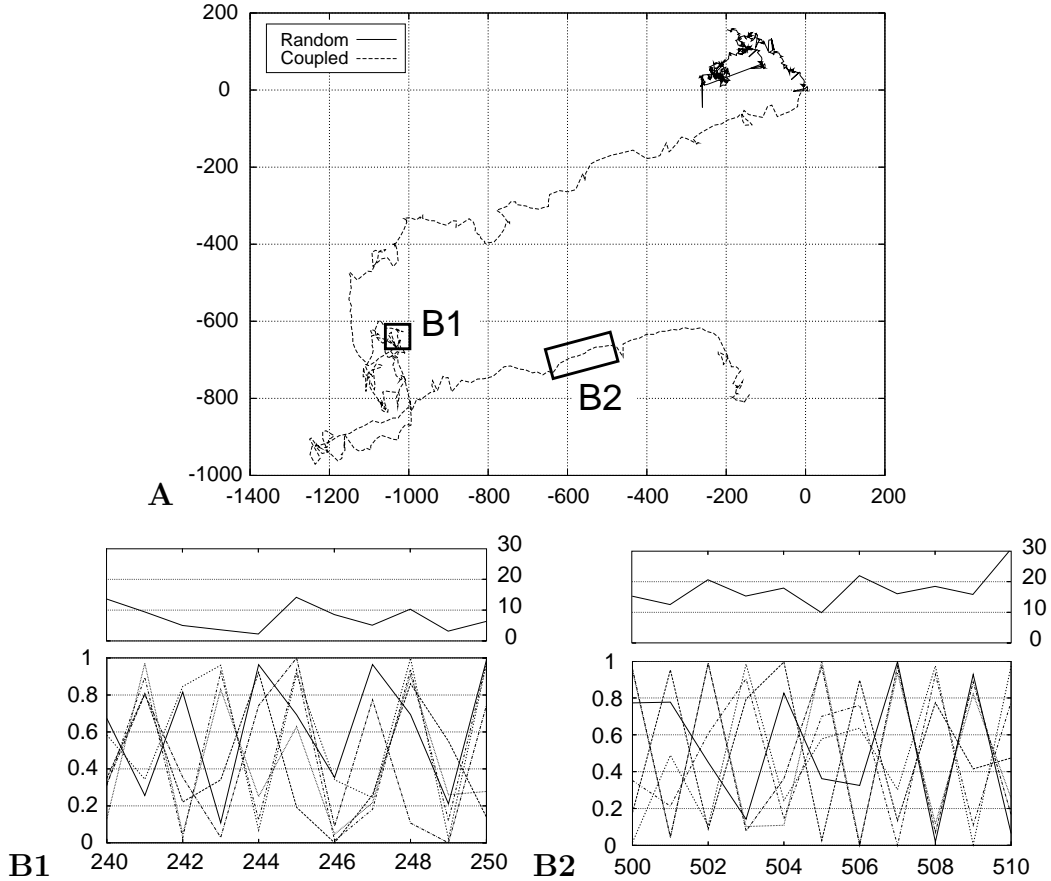


Fig. 8. Trajectories of robot migration. The trajectories for randomly driven legs (solid line) and legs driven by coupled oscillators (dashed line) are compared for the same number (579) of steps (A). The distance from the start position of the robot is shown on the x- and y- axis in cm. Panels B1 and B2 show the migration distance in mm for each time step (top) and the phases of the legs (bottom). The two panels correspond to the movement indicated by the labeled boxes in (A).

where  $x_i, x_j$  correspond to the six van der Pol oscillators,  $\varepsilon, \beta$  represent the nonlinearity, and  $\kappa_{ij}, \sigma_{ij}$  are the coupling constants among the oscillators. In addition, to account for the variability seen in the living oscillators,  $\kappa_{ij}$  and  $\sigma_{ij}$  are slightly varied at every time step with  $\kappa_{ij}^{t+1} = \kappa_{ij}^t + \Delta\kappa_{ij}$  and  $\sigma_{ij}^{t+1} = \sigma_{ij}^t + \Delta\sigma_{ij}$ , where  $\Delta\kappa_{ij}, \Delta\sigma_{ij}$  are Gaussians with zero mean and unit variance. Consequently, as is the case in the *Plasmodium* circuit, the network of oscillators dynamically changes and is never trapped in a stable or periodic state.

A typical trajectory of the robot under control of the van der Pol oscillator system is depicted in figure 8A. Control experiments conducted with in-phase periodic movements of the legs (not shown) and random movements of the legs (solid line in figure 8A) confirmed that the direction of locomotion of the robot is not biased by unintentional asymmetries arising from variations in the construction of the robot.

In the experiment shown the parameters are restricted to the range of:  $\epsilon \in [0, 3]$ ,  $\kappa \in [0, 0.15]$ ,  $\sigma \in [0, 0.2]$  and for simplicity  $\beta = 0$ . Under these conditions we observe both wiggle (figure 8B1) and walking. Directed movement mainly arises from anti-phase oscillation between an oscillator and its opposite oscillator. In particular two adjacent pairs of oscillators that oscillate in anti-phase with their opposite oscillators, but in-phase with each other yield an efficient gait with a stepping movement (figure 8B2).

## 6 Conclusions and Outlook

We have presented an architecture for a cell based robot controller and have shown that cells of *Physarum polycephalum* can provide the functionality required by our design. Previous reports on *Physarum* based oscillator systems used a fixed coupling strength which was determined by the width of the channel connecting oscillators. Our experiments show that light signals can be used to dynamically alter the coupling among oscillators and thus point to a path for interfacing the sensors of the robot with the plasmodium. Clearly, this work is at an early stage and further studies are needed to explore the properties, capabilities and limitations of such a system.

We believe that the use of biological cells for the control of autonomous robots is attractive because of several features difficult to obtain with other technologies. The nano-architecture of the robot controller self-reproduces and is therefore of low cost. Power requirements are exceedingly low; circuits of the type employed here can operate for several days without supply. The controller can be stored for a long time in a dry dormant state and activated by moisture. Moreover, the nano-architecture of the controller comes with a built in self-repair capability. In the foreseeable future it may be possible to use the tools of synthetic biology to customise cell-based robot controllers for a particular application (Blake and Isaacs, 2004; Simpson, 2004). We speculate that cellular controllers are less likely than conventional designs to get stuck in contradictory or undefined situations.

## Acknowledgments

The authors thank Masashi Aono for discussions and technical advice. The work reported here was supported in part by the the Science and Technology Agency of Japan through the Center Of Excellence (COE) program.

## References

- Adamatzky, A., de Lacy Costello, B., Melliush, C., Ratcliffe, N., 2004. Experimental implementation of mobile robot taxis with onboard Belousov-Zhabotinsky chemical medium. *Materials Science & Engineering C* 24 (4), 541–548.
- Blake, W. J., Isaacs, F. J., 2004. Synthetic biology evolves. *Trends in Biotechnology* 22 (7), 321–324.
- Brooks, R., 1990. Elephants don't play chess. *Robotics and Autonomous Systems* 6 (1-2), 3–16.
- Brooks, R., 1991. Intelligence without representations. *Artificial Intelligence* 47, 139–160.
- Cariani, P., 1992. Some epistemological implications of devices which construct their own sensors and effectors. In: Varela, F. J., Bourgine, P. (Eds.), *Toward a Practice of Autonomous Systems: Proceedings of the First European Conference on Artificial Life*. MIT Press, Cambridge, MA, pp. 484–493.
- Conrad, M., 1972. Information processing in molecular systems. *Currents in Modern Biology* (now *BioSystems*) 5, 1–14.
- Conrad, M., 1989. The brain-machine disanalogy. *BioSystems* 22, 197–213.
- Conrad, M., 1995. Scaling of efficiency in programmable and non-programmable systems. *BioSystems* 35, 161–166.
- Conrad, M., 1996. Cross-scale information processing in evolution, development and intelligence. *BioSystems* 38, 97–109.
- Conrad, M., 1999. Molecular and evolutionary computation: the tug of war between context freedom and context sensitivity. *BioSystems* 52, 99–110.
- Durham, A. C. H., Ridgway, E. B., 1976. Control of chemotaxis in *physarum polycephalum*. *The Journal of Cell Biology* 69, 218–223.
- Fujita, M., Kuroki, Y., Ishida, T., Doi, T., 2003. A small humanoid robot sdr-4x for entertainment applications. In: *Proceedings of international conference on advanced intelligent mechatronics*. pp. 435–442.
- Liberman, E. A., 1979. Analog-digital molecular cell computer. *BioSystems* 11, 111–124.
- McCarthy, J., Hayes, P., 1969. Some philosophical problems from the standpoint of artificial intelligence. *Machine Intelligence* 4, 463–502.
- Nakagaki, T., Ueda, T., 1996. Phase switching of rhythmic contraction in relation to regulation of amoeboid behavior by the plasmodium of *physarum polycephalum*. *Journal of Theoretical Biology* 179, 261–267.
- Nakagaki, T., Yamada, H., Toth, A., 2000. Intelligence: Maze-solving by an amoeboid organism. *Nature* 407, 470.
- Nakagaki, T., Yamada, H., Toth, A., 2001. Path finding by tube morphogenesis in an amoeboid organism. *Biophysical Chemistry* 92, 47–52.
- Nakagaki, T., Yamada, H., Ueda, T., 1999. Modulation of cellular rhythm and photoavoidance by oscillatory irradiation in the *physarum* plasmodium. *Biophysical Chemistry* 82, 23–28.
- Nomura, S., 2001. Symbolization of an object and its freedom in biological

- systems. Ph.D. thesis, Kobe University.
- Rambidi, N. G., 2005. Biologically inspired information processing technologies: Reaction-diffusion paradigm. *Int. J. Unconventional Computing* 1 (2), 101–121.
- Renzel, S., Esselborn, S., Sauer, H. W., Hildebrandt, A., 2000. Calcium and malate are sporulation-promoting factors of *physarum polycephalum*. *Journal of Bacteriology* 182 (24), 6900–6905.
- Simpson, M. L., 2004. Rewiring the cell: synthetic biology moves towards higher functional complexity. *Trends in Biotechnology* 22 (11), 555–557.
- Takahashi, K., Uchida, G., Hu, Z., Tsuchiya, Y., 1997. Entrainment of the self-sustained oscillation in a *physarum polycephalum* strand as a one-dimensionally coupled oscillator system. *Journal of Theoretical Biology* 184, 105–110.
- Takamatsu, A., Fujii, T., 2002. Construction of a living coupled oscillator system of plasmodial slime mold by a microfabricated structure. *Sensors Update* 10 (1), 33–46.
- Takamatsu, A., Fujii, T., Endo, I., 2000a. Control of interaction strength in a network of the true slime mold by a microfabricated structure. *BioSystems* 55, 33–38.
- Takamatsu, A., Fujii, T., Yokota, H., Hosokawa, K., Higuchi, T., Endo, I., 2000b. Controlling the geometry and the coupling strength of the oscillator system in plasmodium of *physarum polycephalum* by microfabricated structure. *Protoplasma* 210, 164–171.
- Takamatsu, A., Takahashi, K., Nagao, M., Tsuchiya, Y., 1997. Frequency coupling model for dynamics of responses to stimuli in plasmodium of *physarum polycephalum*. *Journal of Physical Society of Japan* 66, 1638–1646.
- Tani, J., Ito, M., Sugita, Y., 2004. Self-organization of distributedly represented multiple behavior schemata in a mirror system. *Neural Networks* 17 (8-9), 1273–1289.
- Tani, J., Nolfi, S., 1999. Learning to perceive the world as articulated: an approach for hierarchical learning in sensory-motor systems. *Neural Networks* 12, 1131–1141.
- Tero, A., Kobayashi, R., Nakagaki, T., 2005. Coupled-oscillator model with a conservation law in rhythmic amoeboid movements of plasmodial slime mould. *Physica D*(In press.).
- Ueda, T., Matsumoto, K., Kobatake, Y., 1986. Spatial and temporal organization of intracellular adenine nucleotides and cyclic nucleotides in relation to rhythmic motility in *physarum plasmodium*. *Experimental Cell Research* 162 (2), 486–494.
- Ueda, T., Mori, Y., Nakagaki, T., Kobatake, Y., 1988. Action spectra for superoxide generation and UV and visible light photoavoidance in plasmodia of *physarum polycephalum*. *Photochemistry and Photobiology* 48, 705–709.
- Ziegler, J., Dittrich, P., Banzhaf, W., 1998. Towards a metabolic robot control system. In: Holcombe, M., Paton, R. (Eds.), *Information Processing in Cells and Tissues*. Plenum Press, New York, pp. 305–317.